

## Is There Any Evidence to Support the Use of Garlic as a Wormer for Dogs and Cats in the UK?

A Knowledge Summary by

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### PICO question

In dogs and cats, is the oral administration of garlic, compared to no treatment, efficacious at preventing or reducing the intestinal worm burden (species found in the UK)?

### Clinical bottom line

No studies were identified that investigated the efficacy of garlic at preventing intestinal worm burden. Garlic reduced egg and/or larvae counts in the worm species studied. However, where measured, egg and larvae count rapidly (2 days) returned to pretreatment levels once dietary garlic was discontinued. None of the studies included adulticidal action as an outcome measure. In the absence of research to demonstrate high levels of adulticidal action against a range of intestinal wormers at therapeutic, non-toxic levels in cats and dogs, clients should be advised that garlic has not been demonstrated to be an effective anthelmintic (either for multiple or single species use) for use in dogs and cats either to prevent or to treat an intestinal worm burden.

### Clinical Scenario

The veterinary nurse is reading through the worming advice being given in a Facebook group that encourages a natural approach to preventative medicine in dogs and cats. She notices that garlic is being recommended quite frequently by some pet owners as an alternative to a conventional anthelmintic and wonders what the evidence base is for this recommendation. She notes that some owners are recommending its routine use to prevent dogs or cats becoming parasitised and others are recommending it for dogs or cats known to have an intestinal worm burden so she includes both aspects in her PICO.

### The evidence

No papers were identified that addressed the use of garlic to prevent dogs and/or cats becoming parasitised by intestinal worm species. Three papers were identified that either fully (Bastidas, 1969; Ronagh et al. 2015) or partially (Andrei et al., 2011) addressed the intestinal worm reduction aspect of the PICO. Two of the studies focused on dogs (Bastidas, 1969; Andrei et al., 2011) and one of the studies focused on cats (Ronagh et al., 2015). Not all species of intestinal worm known to parasitise cats and/or dogs in the UK were represented, with tapeworm species being the notable exception. All three studies were clinical trials that either used the animal as its own control (Andrei et al., 2011, Bastidas, 1969) or allocated the animals to separate treatment groups (Ronagh et al., 2015). Random allocation of the latter was not reported. Despite being clinical trials, all three studies are very limited with poor data handling and insufficient reporting of the methodology and/or results.

The Andrei et al. (2011) study used the garlic in conjunction with other herbs so any potential anthelmintic effect of garlic is totally confounded with the other components (n = 6, plus water) of the tincture and pumpkin oil preparation used. However, this tincture was associated with a greater than 90% reduction in eggs per gram of faeces for all species (*Toxocara canis*, *Ancylostoma* spp., *Trichocephalus* spp.), with similar findings across both populations (shelter dogs: n = 37, owned dogs: n = 10) studied. Bastidas (1969), with a sample size of one, found that larvae count of *Ancylostoma Caninum* decreased during daily dosing with garlic, but eggs per gram of faeces was only slightly reduced. Rapid recovery to pre-dosing levels (2 days) was observed

following treatment cessation. Finally, Ronagh et al. (2015) found that cats dosed with garlic (n = 5) showed a numerical reduction in *Toxocara cati* eggs on a faecal egg count and a numerical reduction in fecundity rate (number of eggs produced by a female adult *Toxocara cati* worm). No such reduction in either parameter was observed for Control cats (n = 5). None of the studies directly studied the effect of garlic as an adulticide and this remained an important practical limitation in the use of these findings.

## Summary of the evidence

Andrei et al. (2011)	
<b>Population:</b>	<p>Two different populations of dogs were used.</p> <p>Group one: unwormed dogs at a Romanian rescue centre.</p> <p>Group two: dogs owned by people living in Timișoara (Romania).</p> <p>Both sexes, different breeds, and an age range of 2 months–4 years were used, but it is not possible to distinguish how this was distributed across the two groups. No further information is provided.</p>
<b>Sample size:</b>	<p>Group one: 37 dogs</p> <p>Group 2: 10 dogs</p>
<b>Intervention details:</b>	<p>The study design was a before/after design, with each dog used as its own control.</p> <p>The intervention was a home prepared herbal tincture (10% solution), plus a dose of cold pressed pumpkin oil. The herbal tincture contained the following herbs:</p> <ul style="list-style-type: none"> <li>- <i>Inula helenium</i></li> <li>- <i>Tanacetum vulgare</i></li> <li>- <i>Thymus serpyllum</i></li> <li>- <i>Artemisia absinthium</i></li> <li>- <i>Allium ursinum</i></li> <li>- <b><i>Allium sativum</i> (garlic)</b></li> </ul> <p>The quantities of each herbal are not stated, and the references cited are for resources reported in the Romanian language.</p> <p>The dose administered of both the tincture and the pumpkin oil was weight dependent, and was given <i>per os</i> twice daily for 5 days.</p> <p>Experimental timeline</p> <p>Day 0: a faecal sample was obtained from each dog.</p> <p>Day 1–13: at some point over these 13 days each dog was given a weight dependent twice daily dose of the tincture plus the pumpkin oil for 5 days. The actual days this was administered on are not reported.</p>

	<p>Day 14: a faecal sample was obtained from each dog.</p> <p>The Willis and McMaster coproscopic method (no reference provided by the authors) was used to undertake the egg count.</p>
<b>Study design:</b>	Non-randomised controlled trial (before/after design)
<b>Outcome studied:</b>	<p>Worm eggs per gram of faeces, split down by species/class. Samples taken before (day 0) and after (day 14) the intervention was applied.</p> <p>A percentage effectiveness score was then calculated:</p> <p><b>Effectiveness (%) = ((EPG day 0 – EPG day 14) ÷ EPG day 0) X 100</b></p> <p>EPG = Eggs per gram</p>
<b>Main findings: (relevant to PICO question):</b>	<p>In summary, for both groups of dogs, the egg count of the three species present was considerably lower following the intervention.</p> <p>The following values show the mean ± standard error of the mean, associated with each group/day/species. Please note the authors do not conduct analytical statistics so there are no p values. They do report a confidence level of 95% but then appear<sup>1</sup> to report the confidence interval as a range, rather than as lower limit and higher limit.</p> <p>Group 1:</p> <p><i>Toxocara canis:</i></p> <ul style="list-style-type: none"> <li>- Day 0: 1180.40 ± 131.19</li> <li>- Day 14: 121.70 ± 28.61</li> <li>- Effectiveness: 92.55%</li> </ul> <p><i>Ancylostoma spp.:</i></p> <ul style="list-style-type: none"> <li>- Day 0: 1212.96 ± 121.41</li> <li>- Day 14: 131.40 ± 28.39</li> <li>- Effectiveness: 91.87%</li> </ul> <p><i>Trichocephalus spp.:</i></p> <ul style="list-style-type: none"> <li>- Day 0: 1011.54 ± 211.01</li> <li>- Day 14: 123.00 ± 43.34</li> <li>- Effectiveness: 91.34%</li> </ul> <p>Group 2:</p> <p><i>Toxocara canis:</i></p> <ul style="list-style-type: none"> <li>- Day 0: 800.00 ± 117.2</li> <li>- Day 14: 50.00 ± 20.41</li> <li>- Effectiveness: 93.86%</li> </ul> <p><i>Ancylostoma spp.:</i></p> <ul style="list-style-type: none"> <li>- Day 0: 810.00 ± 182.62</li> <li>- Day 14: 70.00 ± 30.00</li> <li>- Effectiveness: 92.57%</li> </ul>

	<p><i>Trichocephalus</i> spp.:</p> <ul style="list-style-type: none"> <li>- Day 0: 508.30 ± 83.08</li> <li>- Day 14: 50.00 ± 18.26</li> <li>- Effectiveness: 93.82%</li> </ul> <p><sup>1</sup>The interpretation made by the author of this Knowledge Summary.</p>
<b>Limitations:</b>	<ul style="list-style-type: none"> <li>- Inadequate reporting of the scientific method (lacks detail, not reproducible given the level of detail reported)</li> <li>- No information regarding the quantity of herbs present in a standardised dose of the tincture</li> <li>- No information as to whether the scientists involved in this study were blinded</li> <li>- Data handling is poor: effect sizes calculated for values that are meaningless, confidence intervals are reported and labeled incorrectly</li> <li>- The author uses parametric measures of central tendency and variation but does not report assessing the distribution of the data and no skew value is reported</li> <li>- Although the tincture appears to have a marked effect on worm burden in dogs it is impossible to quantify the effect, if any, of the addition of garlic to this preparation</li> <li>- The sample size for treatment group two is small</li> </ul>
<b>Bastidas (1969)</b>	
<b>Population:</b>	<p>A dog weighing 10 kg and naturally infected with <i>Ancylostoma caninum</i></p> <p>No further information is available about the dog</p>
<b>Sample size:</b>	One
<b>Intervention details:</b>	<p>The intervention was the addition of garlic to the dog's diet (ground meat, once daily). The dose varied daily (see below).</p> <p>Experimental time line:</p> <p>Day 1–5: Before phase. Dog was fed ground meat only.</p> <p>Day 6–10: During phase. Dog was fed ground meat plus garlic.</p> <p>Day 11–12: After phase. Dog was fed ground meat only.</p> <p>A faecal sample was collected on all 12 days, and a daily egg count (4 replicates, 50 mg samples) undertaken using the Kato method (Martin and Beaver, 1968), and a culture (five replicates, 300 mg samples) per day were made and allowed to stand for 10 days (in a darkened room, at an ambient temperature of 21–23°C). The larvae were then killed using iodine and counted.</p>

	<p>The quantity of garlic fed/ingested was:</p> <ul style="list-style-type: none"> <li>- Day 6: 6.7 g</li> <li>- Day 7: 8.7 g</li> <li>- Day 8: 10.1 g</li> <li>- Day 9: 10.4 g</li> <li>- Day 10: 10.0 g</li> </ul>																																																				
<p><b>Study design:</b></p>	<p>Non-randomised controlled trial (before intervention/during intervention/after intervention)</p>																																																				
<p><b>Outcome studied:</b></p>	<p>There were two outcome measures:</p> <ol style="list-style-type: none"> <li>1. Daily mean egg count (eggs/smear)</li> <li>2. Daily mean larvae count (larvae/culture)</li> </ol>																																																				
<p><b>Main findings: (relevant to PICO question):</b></p>	<p>The author of the research paper reports only raw daily values in a table. With a sample size of one, there are no analytical statistics.</p> <p>To aid visualisation of the results, these raw values have been reported using line graphs created by the author of this Knowledge Summary.</p> <div data-bbox="603 913 1426 1299" data-label="Figure"> <table border="1"> <caption>Data for Figure 1: Mean daily egg count per smear</caption> <thead> <tr> <th>Day of trial</th> <th>Mean daily egg count per smear</th> </tr> </thead> <tbody> <tr><td>Day 1</td><td>370</td></tr> <tr><td>Day 2</td><td>380</td></tr> <tr><td>Day 3</td><td>300</td></tr> <tr><td>Day 4</td><td>330</td></tr> <tr><td>Day 5</td><td>340</td></tr> <tr><td>Day 6</td><td>310</td></tr> <tr><td>Day 7</td><td>280</td></tr> <tr><td>Day 8</td><td>300</td></tr> <tr><td>Day 9</td><td>290</td></tr> <tr><td>Day 10</td><td>290</td></tr> <tr><td>Day 11</td><td>350</td></tr> <tr><td>Day 12</td><td>330</td></tr> </tbody> </table> </div> <p>Figure 1: Mean daily egg count. Nb. before phase/treatment (day 1–5); during phase/treatment (day 6–10); after phase/treatment (day 11–12)</p> <div data-bbox="603 1473 1426 1868" data-label="Figure"> <table border="1"> <caption>Data for Figure 2: Mean daily larvae count per culture</caption> <thead> <tr> <th>Day of trial</th> <th>Mean daily larvae count per culture</th> </tr> </thead> <tbody> <tr><td>Day 1</td><td>1100</td></tr> <tr><td>Day 2</td><td>900</td></tr> <tr><td>Day 3</td><td>800</td></tr> <tr><td>Day 4</td><td>850</td></tr> <tr><td>Day 5</td><td>880</td></tr> <tr><td>Day 6</td><td>750</td></tr> <tr><td>Day 7</td><td>300</td></tr> <tr><td>Day 8</td><td>250</td></tr> <tr><td>Day 9</td><td>200</td></tr> <tr><td>Day 10</td><td>180</td></tr> <tr><td>Day 11</td><td>600</td></tr> <tr><td>Day 12</td><td>880</td></tr> </tbody> </table> </div> <p>Figure 2: Mean daily larvae count. Nb. before phase/treatment (day 1–5); during phase/treatment (day 6–10); after phase/treatment</p>	Day of trial	Mean daily egg count per smear	Day 1	370	Day 2	380	Day 3	300	Day 4	330	Day 5	340	Day 6	310	Day 7	280	Day 8	300	Day 9	290	Day 10	290	Day 11	350	Day 12	330	Day of trial	Mean daily larvae count per culture	Day 1	1100	Day 2	900	Day 3	800	Day 4	850	Day 5	880	Day 6	750	Day 7	300	Day 8	250	Day 9	200	Day 10	180	Day 11	600	Day 12	880
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	<p>(day 11–12)</p> <p>The author of this Knowledge Summary also undertook the following analysis on the data reported in the original paper:</p> <p>Using the formula reported in the Andrei et al. (2011) study, the percentage of effectiveness was calculated. The mean value before feeding garlic (day 1–5) was used as the baseline measurement (EPG day 0), and day 10 (last day that garlic was fed) as the comparator (EPG day 10). The findings were as follows:</p> <ul style="list-style-type: none"> <li>- Effectiveness at reducing egg count by day 10: 14.75% effective<sup>1</sup></li> <li>- Effectiveness at reducing larvae count by day 10: 81.77% effective<sup>2</sup></li> </ul> <p><sup>1</sup> 14.75% = ((342.7 – 296)/342.7) x 100%</p> <p><sup>2</sup> 81.77% = ((927.2)/169)/927.2) x 100%</p>
<b>Limitations:</b>	<ul style="list-style-type: none"> <li>- There was a sample size of one</li> <li>- There is no evidence that blinding was undertaken so the person handling the faecal sample was potentially aware of whether the sample corresponded to days on which garlic was administered</li> <li>- The dose of garlic was not standardised per day (it is not clear whether this is because the quantity offered was different each day, or the quantity voluntarily consumed was different despite offering a fixed amount)</li> <li>- The study findings could have been strengthened by a repeated measures design on this one subject, to help rule out stochastic or alternative explanations for the findings</li> <li>- Limited data handling</li> </ul>
<b>Ronagh et al. (2015)</b>	
<b>Population:</b>	Stray domestic shorthair female cats (weighing circa 3 kg), captured from the streets of Tehran and naturally infected with <i>Toxocara cati</i> roundworms (based on faecal egg counts).
<b>Sample size:</b>	25 cats (five per treatment group), drawn from a wider trapped sample of 100 cats. The 25 cats with the heaviest worm burden were selected from this larger sample.
<b>Intervention details:</b>	<p>Cats were allocated to one of five treatment groups:</p> <ol style="list-style-type: none"> <li>1. Control (no treatment)</li> <li>2. Garlic (fed one 1.25 g garlic tablet daily)</li> <li>3. Black seed (fed 12 g of black seed powder daily)</li> <li>4. Pumpkin (fed 3 g of pumpkin seed powder daily)</li> <li>5. Cloves (fed 6 g of clove powder daily)</li> </ol> <p>Cats were trapped, housed separately, and given 3 days to acclimatise to their environment before the treatments were</p>

	<p>applied.</p> <p>Experimental timeline:</p> <ol style="list-style-type: none"> <li>1. Day 0 (prior to treatment): faeces were collected, and stored in a 10% formalin buffer solution to prevent decay. An egg count was performed at some point subsequent to this.</li> <li>2. Day 7: cats were fed a standard feed ration (not defined in more detail by the authors), either without a supplement (Control group) or with a supplement (experimental treatment groups). The supplement given is outlined in the treatment group description above.</li> <li>3. 7 days after treatments were applied (not clear whether this is 7 days after the start or end of the supplement phase): faeces were collected, and stored in a 10% formalin buffer solution to prevent decay. An egg count was performed (formalin-ether sediment method, no reference provided by the authors) at some point subsequent to this.</li> <li>4. Cats were then euthanised, and the stomach and intestines removed and preserved in 10% formalin. Sections of stomach, duodenum, jejunum and ileum were then stained (haematoxylin and eosin), and examined microscopically for damage). The number of intestinal adult female <i>Toxocara cati</i> worms was counted.</li> </ol>
<p><b>Study design:</b></p>	<p>Controlled trial (not clear if randomised or nonrandomised)</p>
<p><b>Outcome studied:</b></p>	<p>Two outcome measures were relevant to the PICO:</p> <ol style="list-style-type: none"> <li>1. Faecal egg count (<i>Toxocara cati</i> eggs)</li> <li>2. Fecundity rate (<i>Toxocara cati</i> worms)</li> </ol> <p>The fecundity rate was calculated using the following equation:</p> <p><b>Fecundity rate = EPG ÷ Number of female adult worms</b></p> <p>EPG = eggs per gram</p>
<p><b>Main findings: (relevant to PICO question):</b></p>	<p>Please note: only the Control group and the Garlic group results are reported here.</p> <ol style="list-style-type: none"> <li>1. Faecal egg count: The mean (± standard deviation) <i>Toxocara cati</i> eggs per gram of faeces were as follows: <ul style="list-style-type: none"> <li>- Day 0: Control group: 9.4 (± 1.1); Garlic group: 19.0 (± 2.2)</li> <li>- Day 7: Control group: 9.0 (± 1.6); Garlic group: 8.8 (± 0.8)</li> </ul> </li> </ol> <p>The authors also report that the Control group versus Garlic group</p>



was significantly different (T-test, P = 0.003).

Nb. They do not make clear what data this T-test was performed with (the raw data for day 0 or day 7, or a comparison between groups after calculating the difference in EPG between day 0 and day 7 for each individual cat. No other statistical analysis is performed (e.g. within treatment differences) on this data.

2. Fecundity rate:

The authors report only raw data (individual animals) for this data. The raw data can be seen in table 1 below.

Table 1: A comparison of the fecundity rate for cats in the control and garlic treatment groups

Treatment group	Animal no.	Day 0: fecundity rate	Day 7: fecundity rate
Control	1	5.5	5.5
	2	5	5
	3	9	9
	4	4.5	4
	5	8	7
Garlic	1	6.66	3
	2	11	5
	3	9	4.5
	4	9.5	4
	5	16	8

Descriptive statistics (below) have been performed on this data set by the Knowledge Summary author using the same parameters used by the paper authors in the first outcome measure. Data distribution or skewness has not been evaluated.

The mean ( $\pm$  standard deviation) fecundity rate was as follows:

- Day 0: Control group: 6.4 ( $\pm$  2.0); Garlic group: 10.4 ( $\pm$  3.5)
- Day 7: Control group: 6.1 ( $\pm$  1.9); Garlic group: 4.9 ( $\pm$  1.9)

**Limitations:**

- The authors do not report how they allocated the cats to the different treatment groups so it is not known whether cats were randomly allocated to receive the different treatments.
- There is no evidence that the researchers or any other personnel involved in the study (if any) were blinded as to the treatments when collecting/analysing data.
- Insufficient experimental detail (in both the methods and results sections) is available to fully understand or interpret the study findings.
- There is insufficient detail in the methods to fully appraise scientific rigour (e.g. in relation to egg count methodology, use of duplicate samples, etc.)
- Statistical analysis is limited and it is unclear what the p

	<p>value reported relates to. The authors appear to have compared treatment groups before treatment application and not after. However it is possible that they have used 'within treatment differences in egg count between day 0 and day 7' data, and then conducted a between treatment group T-test on the 'differences' data. This would allow them to undertake one statistic test to compare between treatment groups, while also handling the data in a manner that would increase the likelihood of finding significant differences where there is a lot of variation between dogs in terms numbers of eggs present in faeces at either time point. The authors do not tell us though that they do this and this lack of detail is a study failing.</p> <ul style="list-style-type: none"> <li>- The authors do not justify their use of parametric statistical analysis or choice of measure of central tendency.</li> <li>- The sample size per treatment group is very small and does not appear to be based on a power calculation or other approach to determine an appropriate sample size.</li> <li>- The treatment groups differed in mean number of eggs per gram of faeces (the Garlic group had approximately double the number of eggs compared to the Control group) on day 0 (prior to treatment application). Likewise, the fecundity rate of the Garlic group was higher than the Control group at the start of the study (day 0).</li> <li>- The authors report that garlic reduced the number of female adult worms and refer the reader to table 2 in demonstration of this. However, table 2 refers to fecundity rate (not the number of adult female worms) and this value cannot be used to quantify number of adult female worms present using the information provided. The number of adult female worms was not listed as a planned outcome measure, and is not reported anywhere else in the paper.</li> </ul>
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## Appraisal, application and reflection

Plant-based anthelmintics have been suggested as a potential alternative to overcome increasing resistance to conventional anthelmintics (French, 2018). The use of garlic as a traditional anthelmintic for dogs with intestinal worms has been reported (n = 1 interviewee) in a study exploring central-southern Italy's ethnoveterinary practices (Guarrera et al., 2008), and more widely elsewhere in pigs (Lans et al., 2007; Bartha et al., 2015) and ruminants (Lans et al., 2007; Bullitta et al., 2018). Thus, the promotion of garlic as an anthelmintic in dogs and cats is probably derived from traditional ethnobotanical medical practices. More recently, there has been some growth in scientific interest in its potential anthelmintic properties in a range of mammalian and avian species. Extracts from garlic bulbs shown to have in vivo (e.g. Palacios-Landin et al., 2015, but see e.g. Worku, 2009; Velkers et al., 2011) and/or in vitro (e.g. Palacios-Landin et al., 2015; Orengo et al., 2016; Tavassoli et al., 2018) efficacy (differing stages of the life cycle, dependent on the study) against various species of helminth. This includes in vitro activity against some species (*Toxocara canis*, *Ancylostoma caninum*) that infest dogs (Orengo et al., 2016). Consequently, scientific and veterinary growth in its interest in a wormer for dogs and cats may be likely to develop over time.

No English language papers were identified that addressed the prevention aspect of the PICO. Three papers were identified that either fully (Bastidas, 1969; Ronagh et al., 2015) or partially (Andrei et al., 2011) addressed the treatment aspect of the PICO. Two of the studies focused on dogs (Bastidas, 1969; Andrei et

al., 2011) and one of the studies focused on cats (Ronagh et al., 2015). A further abstract (Bekirov et al., 1979) that examined the use of garlic (in conjunction with other ingredients) as a canine anthelmintic effective against *Echinococcus* or *Taenia hydatigena* was excluded as the main paper was in Russian but reported 92–94.8% and 100% efficacy respectively against each species. The relative lack of studies that addressed the PICO also meant that some of the intestinal worms known to affect dogs and cats in the UK did not have any evidence available to address the use of garlic as an anthelmintic for that species. Studies focusing on tapeworm species were notably absent, with the exception of the Bekirov study, which combined garlic with several other products thereby confounding interpretation of the efficacy of garlic per se as an anthelmintic. Thus, any positive anthelmintic effects at the level of the individual species may still limit clinical use to the practitioner or owner when seeking an anthelmintic effective against a broad range of intestinal worms.

All three studies included in this Knowledge Summary were clinical trials, which either used the animal as its own control (Andrei et al., 2011, Bastidas, 1969) or allocated the animals to separate treatment groups (Ronagh et al., 2015). However, despite being clinical trials, all of the studies showed clear limitations in terms of methodological approach and/or study methodology reporting and/or results reporting and highlight the importance of not using the evidence pyramid (see: O'Connor, 2017 for a discussion on the limitations to the evidence pyramid) in isolation when evaluating the relative quality of a study. Furthermore, the outcome measures used by each of the studies used are unlikely to address the clinical need of veterinary practitioners or clients seeking an anthelmintic that will kill intestinal worms present at the point of dosing the dog or cat.

In the Andrei et al. (2011) study a 90% reduction in eggs per gram of faeces for all species (*Toxocara canis*, *Ancylostoma* spp, *Trichocephalus* spp.) following a twice daily weight dependent dose of their worming preparation (tincture and pumpkin oil). Similar results were obtained for both populations (shelter dogs: n = 37, owned dogs: n = 10) studied. However, this worming preparation used the garlic in conjunction with other herbs so any potential anthelmintic effect of garlic is totally confounded with the other components (n = 6, plus water) of the tincture and pumpkin oil preparation used. Thus, it is impossible to quantify the relative contribution (positive, negative, additive, synergistic, or no effect at all) of garlic to these findings. In defence of the authors, this study was designed to test the efficacy of this worming preparation rather than to investigate the efficacy of garlic in isolation as an anthelmintic. However, this study is also problematic in terms of its scientific quality, with authors failing to report tincture composition in sufficient detail, with no detail available on the quantity of each herb added to the tincture preparation. Dosing standardisation was achieved through product dosing based on the weight of the dog, but this is only described in terms of quantity of the tincture plus pumpkin oil supplied. Despite the most impressive sample size (relative to the other two studies reported here), the authors do not perform analytical statistics on their findings, and while they describe reporting the confidence intervals (which can be used in preference to p values), they appear to be reporting this as one value rather than as an upper and lower limit which limits its value in interpreting the data. However, the descriptive statistics do suggest that the before and after treatment faecal egg counts would be significantly different across all three of the worm species studied (and the direction of the effect is similar for both shelter and owned dogs) should a suitable analytical test be performed. Despite this, the study suffers from another key issue when considering the clinical application of this tincture, and that is that the outcome measure assessed did not include either a direct or indirect (proxy) measurement of the effect of the preparation on adult worm mortality and/or long-term fecundity. The study finished immediately after the end of the tincture and pumpkin oil dosing period. Thus, all that is known is that this worming preparation had effects on egg production during the period of dosing, without anything to indicate the possible reason for this reduction. This is a clinically important issue that is of relevance to any anthelmintic product selection, and represents a major study limitation within the context of any clinician considering using this worming preparation in preference to any product with known adulticidal efficacy.

The second of the studies evaluated (Bastidas, 1969) was included as a clinical trial based on its study methodology (before, during, after treatment) allowing it to meet the inclusion criteria but it had a sample size of one dog, and with each study phase undertaken only once, findings were potentially explicable, either partially or fully, by other undefined or unreported effects. This should be borne in mind when considering the reported findings. This study found that larvae count of *Ancylostoma caninum* decreased during daily

dosing with garlic (non-standardised dose), but eggs per gram of faeces remained similar following a five-day dosing period. Application of the Andrei *et al.* (2011) equation for evaluating anthelmintic efficacy to Bastidas' (1969) raw data indicated that efficacy at reducing egg count after 5 days of garlic treatment was only 14.75%. This was much lower than the Andrei *et al.* (2011) study, and suggests that other components of the Andrei *et al.* study's worming preparation may have explained the increased efficacy at reducing egg count identified in that study. However, there are other differences in the study methodology and lack of detail regarding the tincture preparation mean that meaningful comparisons are difficult to draw. Garlic appeared more effective at reducing larvae count and was 81.77% effective at reducing larvae count by day 5 (the last day) of treatment. However, it is important to note that this effect was very short lived and mean larvae count increased rapidly (1 day) following discontinuation of the garlic and returned to approximately pretreatment levels only 2 days after discontinuation of the garlic. Again, while the presence of viable adult female worms was not an outcome measure of this study, these post-treatment changes in larvae count suggest that the addition of garlic to the diet at this dosage and dosing period did not affect adult female worm mortality or longer-term fecundity rates.

Finally, Ronagh *et al.* (2015) found that cats dosed with garlic ( $n = 5$ ) showed a numerical reduction in *Toxocara cati* eggs on a faecal egg count and a numerical reduction in fecundity rate (number of eggs produced by a female adult *Toxocara cati* worm). No such reduction in either parameter was observed for Control cats ( $n = 5$ ). However, this study euthanised the cats at the end of the study (to assess fecundity rate and gastrointestinal damage to the mucosa) and did not measure faecal egg counts for a few days post-treatment cessation. Thus, while it is known that egg counts were lower, and this reduction was probably due to a reduction in the number of eggs produced by each viable female, it is not known whether any inhibitory effect of the garlic is temporary (i.e. females will increase egg production when the garlic is discontinued) or whether it is more permanent (e.g. through increased morbidity/mortality rates of adult female worms). In the light of the Bastidas (1969) study findings this is an important consideration. This study did count the number of adult female worms present within the intestines at the point of euthanasia of both the Control group and the Garlic group but does not report this information. However, the authors do not report how the cats were allocated to their respective treatment groups. Frustratingly, the pretreatment faecal egg count demonstrates that the Control cats had a lower mean ( $\pm$  standard deviation) faecal egg count ( $9.4 \pm 1.1$ ) and fecundity rate ( $6.4 \pm 2.0$ ) than the Garlic group (egg count:  $19.0 \pm 2.0$ ; fecundity rate:  $10.4 \pm 3.5$ ), with important implications for data handling, analysis and interpretation. The authors' report a significant effect of treatment group (Garlic versus Control) but fail to report what data was analysed to obtain this probability value, and its value to the data interpretation is thereby questionable. With better management of subject allocation to the treatment groups (for example by using faecal egg counts to rank cats according to worm burden severity and then allocating to treatments using a randomised block approach) this study could have been strengthened. It is not clear why this was not undertaken as the authors originally trapped 100 cats, and retained the 25 most *Toxocara cati* parasitised cats to use in this study, so this limitation could have been addressed at the study outset.

In summary, based on the limited and relatively poor quality studies available to address the PICO, garlic may have a temporary inhibitory action on larvae and/or egg production of the intestinal worm species studied but none of the studies directly investigated the effect of garlic on adult worm mortality or viability. However, where a proxy measure was used (egg/larvae production after treatment cessation) this suggested that garlic did not have adulticidal action against *Ancylostoma caninum*. In the absence of research to demonstrate high levels of adulticidal action against a range of intestinal wormers at therapeutic, non-toxic levels in cats and dogs, clients should be advised that garlic is not proven as an effective anthelmintic (either against multiple species or a single species) for use in dogs and cats with to prevent, or to treat, an intestinal worm burden.

Search Strategy	
Databases searched and dates covered:	Pubmed, accessed via the NCBI website (01/01/1900 – 20/02/2019); Web of Science (1990 – 20/02/2019)
	<p>Pubmed &amp; Web of Science search:</p> <p>(dog OR dogs OR canine OR canid OR canis OR bitch OR bitches OR pup OR puppy OR puppies OR cat OR cats OR feline OR felid OR kitten OR kittens) AND (garlic OR “allium sativum”) AND (worm OR tapeworm OR tape-worm OR “tape worm” OR roundworm OR round-worm OR “round worm” OR hookworm OR hook-worm OR “hook worm” OR whipworm OR whip-worm OR “whip worm” OR flatworm OR “flat worm” OR flat-worm OR endoparasite OR endo-parasite OR parasite OR parasitic OR anthelmintic OR ascarid OR ascaris OR larvae OR toxocara OR toxascaris OR ancylostoma OR trichuris OR uncinaria OR Dipylidium OR Taenia OR echinococcus OR cestode OR cestodes OR nematode OR nematodes OR Trematode OR Trematodes OR Fluke OR Flukes OR Nanophytus OR heterophyes OR cryptocotyle OR apophallus OR alaria)</p> <p>CAB Abstract search:</p> <ol style="list-style-type: none"> <li>(dog or dogs or canine or canid or canis or bitch or bitches or pup or puppy or puppies or cat or cats or feline or felid or kitten or kittens).mp. [mp=abstract, title, original title, broad terms, heading words, identifiers, cabicodes]</li> <li>(garlic or allium sativum).mp. [mp=abstract, title, original title, broad terms, heading words, identifiers, cabicodes]</li> <li>(worm or tapeworm or tape-worm or tape worm or roundworm or round-worm or round worm or hookworm or hook-worm or hook worm or whipworm or whip-worm or whip worm or flatworm or flat worm or flat-worm or endoparasite or endo-parasite or parasite or parasitic or anthelmintic or ascarid or ascaris or larvae or toxocara or toxascaris or ancylostoma or trichuris or uncinaria or Dipylidium or Taenia or echinococcus or cestode or cestodes or nematode or nematodes or Trematode or Trematodes or Fluke or Flukes or Nanophytus or heterophyes or cryptocotyle or apophallus or alaria).mp. [mp=abstract, title, original title, broad terms, heading words, identifiers, cabicodes]</li> <li>1 and 2 and 3</li> </ol>
Dates searches performed:	Pubmed (20/02/2019); Web of Science (20/02/2019); CAB Abstracts (20/02/2019)

Exclusion / Inclusion Criteria	
Exclusion:	Pre-defined exclusion criteria: non-English language, popular press articles, in vitro studies, conference abstracts
Inclusion:	Any comparative study in which the effect of garlic on intestinal worms in dogs or cats was studied

Search Outcome						
Database	Number of results	Excluded – did not answer the PICO question	Excluded – not English language	Excluded – conference abstract only	Excluded – duplicates	Total relevant papers
Pubmed	8	7	0	0	0	1
Web of Science	10	0	0	0	0	0
CAB Abstracts	17	14	1	0	0	2
Total relevant papers when duplicates removed						3

## CONFLICT OF INTEREST

The author declares no conflicts of interest.

## REFERENCES

1. Andrei, S., Ilie, M. S., Mederle, N. & Darabus, G. (2011) Testing the effectiveness of a plant extract in the therapy on some endoparasites in dogs. *Lucrari Stiintifice - Medicina Veterinara, Universitatea de Stiinte Agricole si Medicina Veterinara "Ion Ionescu de la Brad" Iasi*, 54(3), pp. 247–254.
2. Bartha, S. G., Quave, C. L., Balogh, L., Papp, N. (2015) Ethnoveterinary practices of Covasna County, Transylvania, Romania. *Journal of Ethnobiology and Ethnomedicine* 11: 35 [online] Available from: <https://ethnobiomed.biomedcentral.com/articles/10.1186/s13002-015-0020-8> (accessed: 16/05/2019) DOI: <http://dx.doi.org/10.1186/s13002-015-0020-8>
3. Bastidas, G. J. (1969) Effect of ingested garlic on *Necator americanus* and *Ancylostoma caninum*. *The American Journal of Tropical Medicine and Hygiene*, 18(6), pp. 920–923. DOI: <http://dx.doi.org/10.4269/ajtmh.1969.18.920>
4. Bekirov, R. E., Azimov, Sh. A., Oripov, A. O., & Dzhumaev, Z. (1979) The efficacy of granules against cestodes in dogs. *Veterinariya, Moscow* 8, pp. 50–51.
5. Bullitta, S., Re, G. A., Manunta, D. I., & Oiluzza, G. (2018) Traditional knowledge about plant, animal,



- and mineral-based remedies to treat cattle, pigs, horses, and other domestic animals in the Mediterranean island of Sardinia. *Journal of Ethnobiology and Ethnomedicine* 14: 50 [online] Available from: <https://ethnobiomed.biomedcentral.com/articles/10.1186/s13002-018-0250-7> (accessed: 16/05/2019) DOI: <http://dx.doi.org/10.1186/s13002-018-0250-7>
6. Buona, F., Pacifico, L., Piantedosi, D., Sgroi, G., Neola, B., Roncoroni, C., Genovese, A., Rufrano, D., & Veneziano, V. (2019) Preliminary Observations of the Effect of Garlic on Egg Shedding in Horses Naturally Infected by Intestinal Strongyles. *Journal of Equine Veterinary Science* 72: 79–83. DOI: <http://dx.doi.org/10.1016/j.jevs.2018.10.025>
  7. French, K. E. (2018) Plant-Based Solutions to Global Livestock Anthelmintic Resistance. *Ethonobiology Letters* 9(2): 110–123. DOI: <http://dx.doi.org/10.14237/ebl.9.2.2018.980>
  8. Guarrera, P. M., Lucchese, F., & Medori, S. (2008) Ethnophytotherapeutical research in the high Molise region (Central-Southern Italy). *Journal of Ethnobiology and Ethnomedicine* 4: 7 [online] Available from: <https://ethnobiomed.biomedcentral.com/articles/10.1186/1746-4269-4-7> (accessed: 16/05/2019) DOI: <http://dx.doi.org/10.1186/1746-4269-4-7>
  9. Lans, C., Turner, N., Khan, T., Brauer, G., & Boepple, W. (2007) Ethnoveterinary medicines used for ruminants in British Columbia, Canada. *Journal of Ethnobiology and Ethnomedicine* 3: 11 [online] Available from: <https://ethnobiomed.biomedcentral.com/articles/10.1186/1746-4269-3-11> (accessed: 16/05/2019) DOI: <http://dx.doi.org/10.1186/1746-4269-3-11>
  10. O'Connor, A. (2017) Is the simplicity of the evidence pyramid actually detrimental for understanding evidence? *Veterinary Evidence* 2(1). DOI: <http://dx.doi.org/10.18849/ve.v2i1.100>
  11. Orengo, K. O., Maitho, T., & Mbaria, J. 2016. In vitro anthelmintic activity of *Allium sativum*, *Allium cepa* and *Jatropha curcas* against *Toxocara canis* and *Ancylostoma caninum*. *African Journal of Pharmacy and Pharmacology* 10(21): 465–471.
  12. Martin, L. K. & Beaver, P. C. (1968) Evaluation of Kato thick-smear technique for quantitative diagnosis of helminth infections. *American Journal of Tropical Medicine and Hygiene* 17, pp. 382–391. DOI: <http://dx.doi.org/10.4269/ajtmh.1968.17.382>
  13. Palacios-Landin, J., Mendoza-de Gives, P., Salinas-Sanchez, D. O., Lopez-Arellano, M. E., Liebano-Hernandez, E., Hernandez-Velazquez, V. M., & Valladares-Cisneros, M. G. (2015) In vitro and in vivo nematocidal activity of *Allium sativum* and *Tagetes erecta* extracts Against *Haemonchus contortus*. *Turkish Journal of Parasitology* 31: 277–282. DOI: <http://dx.doi.org/10.5152/tpd.2015.4523>
  14. Ronagh, K., Gharouni, A., Bahadori, S. R., Zakian, A., Gholami, N., Rezaeian, H. & Shahraki, M. S. (2015) Effect of *Nigella sativa*, *Allium sativum*, *Syzygium aromaticum* and *Cucurbita maxima* on *Toxocara cati* fecal egg count in stray cats. *Online Journal of Veterinary Research*, 19(5), pp. 325–330.
  15. Tavassoli, M., Jalilzadeh-Amin, G., Fard, V. R. B., & Esfandiarpour, R. 2018. The in vitro effect of *Ferula asafoetida* and *Allium sativum* extracts on *Strongylus* spp. *Annals of Parasitology* 64(1): 59–63.
  16. Velkers, F. C., Dieho, K., Pecher, F. W. M., Vernooij, J. C. M., van Eck, J. H. H., & Landman, W. J. M. (2011) Efficacy of allicin from garlic against *Ascaridia galli* infection in chickens. *Poultry Science* 90(2): 364–368. DOI: <http://dx.doi.org/10.3382/ps.2010-01090>
  17. Worku, M., Franco, R., & Baldwin, K. (2009) Efficacy of garlic as an anthelmintic in adult Boer goats. *Archives of Biological Sciences* 61(1): 135–140. DOI: <http://dx.doi.org/10.2298/ABS0901135W>

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